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INTRODUCTION

The 26S proteasome is the proteolytic machine of the ubiquitin-proteasome system. This pathway is of particular importance since it is involved in the degradation of most intracellular proteins. Major biological processes, such as cell cycle progression, apoptosis, DNA repair, epitope generation and cell quality control are tightly regulated by this system. Many studies have demonstrated that a dysregulation of this machinery is related to various pathologies such as cancers and the proteasome has recently been identified as a pharmacological target for their treatment.

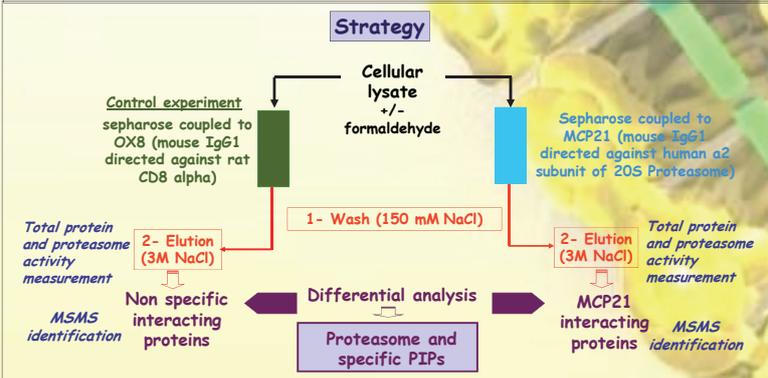
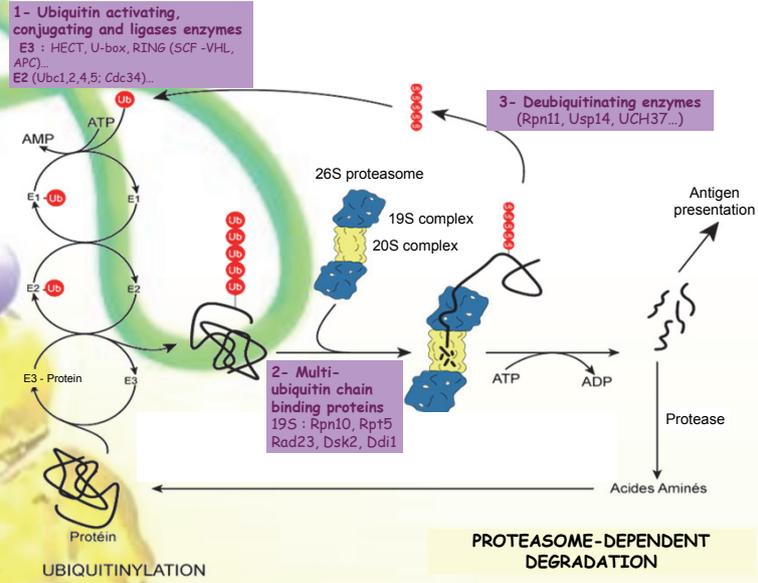
The 26S proteasome is a 2.4 MDa complex composed of multisubunit subcomplexes : a core protease, the 20S proteasome, and two regulatory elements, the 19S particles. A variety of proteins, named "Proteasome Interacting Proteins" (PIPs), interact with the proteasome.

Despite its physiological importance, many aspects of the mammalian proteasome structural organization and regulation remain to be understood. It is known however that its subunit composition and dynamic association to various proteins regulate its stability and activity upon diverse stimuli.

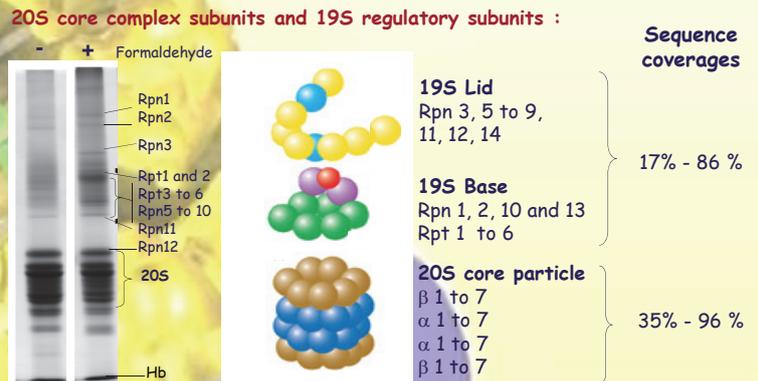
Therefore, we developed a new affinity purification strategy to characterize all proteasome complexes and PIPs in human erythrocytes (Bousquet-Dubouch *et al.*, 2009). This new single-step procedure, based on the high-affinity binding of a subunit of the 20S core particle to a monoclonal antibody, permits to detect endogenous interactions without relying on over-expression or tagging strategies and can be used with whatever human sample as starting material.

Subsequent proteomic analyses identified all proteasomal subunits, known regulators and recently assigned partners. Moreover, other proteins implicated at different levels of the ubiquitin-proteasome system were also identified for the first time.

This novel approach, through the identification of partners affecting proteasomal function, will help to better understand the function of this complex proteolytic machine in different cell physiological states.



Identified proteins in purified proteasome complexes



Other regulatory complex subunits : PA28 (α/β) and PA200 16% - 88%
 Our strategy enables to purify 20S proteasome and its known regulators with high recoveries

Specific human erythrocytes Proteasome Interacting Proteins (PIPs) belonging to the Ubiquitin Proteasome System

Description	Fonction
Usp14, UCH37, USP7 ^{2,3} (HAUSP)	Deubiquitinating enzymes
SCF [Skp1, Cul1 ² , Fbox7, Rbx1 ²], E3C ² , Cullin-associated Nedd8-dissociating protein ^{1,3}	E3 ligases or related proteins
PACs 1, 2 and 3 ^{1,3}	Chaperones involved in proteasome assembly
VCP protein ³ , HSPs (70, 90 ¹), HSC71, HSPA5 ¹	Other chaperones
Ecm29 ¹ , HSP90 ¹	Proteins involved in proteasome stability
PI31	Proteasome inhibitor
hHR23B ¹ (UBA/UBL), eEF1α ¹ , Ddi1 ¹	Substrate delivery to the proteasome
Poly-Ubiquitin (Lys48-linked)	Major signal for protein degradation by the proteasome

1: Identified only after formaldehyde cross-linking
 2: Identified only without formaldehyde cross-linking
 3: New PIPs
 Hausp, a deubiquitinase involved in the stability of p53 was further validated using a reverse immuno-purification experiment

Benefit of using Formaldehyde crosslink

The spectral counting approach (Liu *et al.*, 2004) was used to estimate the relative abundance of each proteasome subunit in the formaldehyde-treated sample versus in the non-treated sample (using MFPaQ software - Bouysssié *et al.*, 2007)

Name	Accession #	Unique peptides #		Sequence coverage (%)		MS/MS counting F/N		STREBY
		N	F	N	F	Average	Ratio	
20S								
α								
α1	P09290	29	32	64	1.0	0.0	0.0	
α2	P21787	30	29	89	0.6	0.0	0.0	
α3	P21788	27	27	88	1.2	0.6	0.0	
α4	O14838	34	85	82	0.9	0.2	0.0	
α5	P21786	10	26	75	0.9	0.2	0.1	
α6	P21786	35	43	95	1.0	0.2	0.0	
α7	P21786	35	41	79	0.9	0.2	0.1	
β1	P28072	15	15	79	1.1	0.2	0.0	
β2	Q29456	18	22	75	1.1	0.9	0.0	
β3	P46720	17	22	65	1.1	0.1	0.0	
β4	P46721	21	27	95	0.8	0.3	0.0	
β5	P28074	25	22	89	0.9	0.2	0.1	
β6	P28076	22	27	81	0.9	1.0	0.1	
β7	P28076	12	12	97	1.0	0.1	0.0	
β8	P28065	6	5	34	1.0	1.1	0.2	
β11	P46726	7	7	37	1.3	0.6	0.0	
β12	P28050	13	15	60	1.1	0.3	0.0	
19S								
Base								
Base1	P33988	88	87	89	1.2	0.2	0.0	
Base2	P33989	89	88	92	1.2	0.2	0.0	
Base3	P42456	56	37	84	0.1	1.5	0.2	
Base4	P28051	42	44	79	1.6	0.9	0.0	
Base5	P28052	29	35	66	1.7	0.2	0.0	
Base6	P28053	12	12	74	1.9	0.1	0.0	
Base7	P28054	66	64	69	1.9	0.4	0.0	
Base8	P28055	36	37	75	1.9	0.1	0.0	
Base9	P28056	34	39	80	1.9	0.1	0.0	
Base10	P28057	34	35	65	0.7	1.6	0.2	
Base11	P28058	34	39	72	1.0	1.2	0.1	
Base12	P28059	33	36	79	1.4	0.3	0.0	
Base13	P28060	17	20	24	2.1	0.2	0.0	
Base14	P28061	25	28	78	0.8	2.5	0.6	
Base15	P28062	14	11	41	1.6	3.2	1.9	
Base16	P28063	13	10	65	0.6	4.9	2.0	
Base17	P28064	12	14	64	0.6	3.2	1.2	
Lid								
Lid1	P28065	4	0	11	1.7	4.0	1.0	
Lid2	Q29384	6	0	17	1.7	4.0	1.0	

Our purification strategy is based on the use of a 20S subunit as a bait. This can explain why *in-vivo* crosslinking with formaldehyde permits to maintain the most remote subunits from the 20S core. Formaldehyde therefore helps to preserve the weak association between the 20S core particle and its regulators.

CONCLUSIONS

- The immuno-purification of proteasome complexes is possible in a single-step procedure without relying on overexpression or tagging strategies and with whatever human sample as starting material.
- In-vivo* formaldehyde cross-linking stabilizes weak interactions between the 20S core particle and its activators.
- Subsequent proteomic analyses identified all proteasomal subunits, known regulators and recently assigned partners. Moreover, other proteins implicated at different levels of the ubiquitin-proteasome system were also identified. Among these, Hausp, a deubiquitinase involved in the stability of p53, was further validated as a new PIP by a reverse immuno-precipitation experiment. The presence of Hausp within proteasome complexes might be a mean to regulate its activity.

REFERENCES

Bousquet-Dubouch *et al.* 2009 MCP 8, 1150-1164
 Liu *et al.* 2004 Anal Chem 76, 4193-201
 Bouysssié *et al.* 2007 MCP 6, 1621-1637